# Package 'wingen'

February 27, 2024

Title Continuous Mapping of Genetic Diversity

Version 2.1.1

**Description** Generate continuous maps of genetic diversity using moving windows with options for rarefaction, interpolation, and masking as described in Bishop et al. (2023) <doi:10.1111/2041-210X.14090>.

**License** MIT + file LICENSE

Encoding UTF-8

LazyData true

RoxygenNote 7.3.1

**Imports** automap, crayon, dplyr, furrr, gdistance, graphics, grDevices, ggplot2, hierfstat, magrittr, pegas, purrr, raster, rlang, sf, terra, tidyr, tidyselect, utils, vcfR, viridis

**Suggests** adegenet, covr, devtools, future, knitr, MASS, rmarkdown, stringr, SpatialKDE, testthat (>= 3.0.0)

VignetteBuilder knitr

**Config/testthat/edition** 3

**Depends** R (>= 3.5.0)

NeedsCompilation no

Author Anusha Bishop [aut, cre] (<https://orcid.org/0000-0003-1731-8683>), Anne Chambers [aut] (<https://orcid.org/0000-0002-7369-0108>), Ian Wang [aut] (<https://orcid.org/0000-0003-2554-9414>)

Maintainer Anusha Bishop <anusha.bishop@berkeley.edu>

**Repository** CRAN

Date/Publication 2024-02-27 16:40:03 UTC

# **R** topics documented:

circle_gd	. 2
circle_general	. 4
coords_to_raster	. 6

get_geodist	7
get_resdist	8
ggplot_count	9
ggplot_gd	10
krig_gd	10
load_middle_earth_ex	12
load_mini_ex	13
lotr_coords	
lotr_lyr	
lotr_range	
lotr_vcf	
mask_gd	
mini_coords	
mini_lyr	
mini_vcf	
mini_vcf_NA	
plot_count	
plot_gd	
preview_gd	
resist_gd	
resist_general	
vcf_to_dosage	
window_gd	
window_general	28
	31
	51

## Index

circle_gd	Create a moving window map of genetic diversity using a circle win-
	dow

# Description

Generate a continuous raster map of genetic diversity using circle moving windows

## Usage

```
circle_gd(
  gen,
  coords,
  lyr,
  maxdist,
  distmat = NULL,
  stat = "pi",
  fact = 0,
  rarify = FALSE,
  rarify_n = 2,
  rarify_nit = 5,
```

# circle\_gd

```
min_n = 2,
fun = mean,
L = "nvariants",
rarify_alleles = TRUE,
sig = 0.05
)
```

gen	genetic data either as an object of type vcf or a path to a vcf file ( <i>note:</i> order matters! The coordinate and genetic data should be in the same order; there are currently no checks for this)
coords	coordinates of samples as sf points, a two-column matrix, or a data.frame repre- senting x and y coordinates (see Details for important information about projec- tions)
lyr	SpatRaster or RasterLayer to slide the window across (see Details for important information about projections)
maxdist	maximum geographic distance used to define neighborhood; any samples further than this distance will not be included (this can be thought of as the neighborhood radius) Can either be (1) a single numeric value or (2) a SpatRaster where each pixel is the maximum distance to be used for that cell on the landscape (must be the same spatial scale as lyr).
distmat	distance matrix output from get_geodist (optional; can be used to save time on distance calculations)
stat	genetic diversity statistic(s) to calculate (see Details, defaults to "pi"). Can be a single statistic or a vector of statistics
fact	aggregation factor to apply to lyr (defaults to 0; <i>note:</i> increasing this value reduces computational time)
rarify	if rarify = TRUE, rarefaction is performed (defaults to FALSE)
rarify_n	if rarify = TRUE, number of points to use for rarefaction (defaults to min_n)
rarify_nit	if rarify = TRUE, number of iterations to use for rarefaction (defaults to 5). Can also be set to "all" to use all possible combinations of samples of size rarify_n within the window.
min_n	minimum number of samples to use in calculations (any focal cell with a window containing less than this number of samples will be assigned a value of NA; defaults to 2)
fun	function to use to summarize rarefaction results (defaults to mean, must take na.rm = TRUE as an argument)
L	for calculating "pi", L argument in pi.dosage function. Return the average nucleotide diversity per nucleotide given the length L of the sequence. The wingen default is $L =$ "nvariants" which sets L to the number of variants in the VCF. If L = NULL, returns the sum over SNPs of nucleotide diversity ( <i>note:</i> L = NULL is the pi.dosage default which wingen does not use)
rarify_alleles	for calculating "biallelic_richness", whether to perform rarefaction of al- lele counts as in allelic.richness (defaults to TRUE)

sig for calculating "hwe", significance threshold (i.e., alpha level) to use for hardyweinberg equilibrium tests (defaults to 0.05)

#### Details

Coordinates and rasters should be in a Euclidean coordinate system (i.e., UTM coordinates) such that raster cell width and height are equal distances. As such, longitude-latitude systems should be transformed before using dist\_gd. Transformation can be performed using st\_set\_crs for coordinates or project for rasters (see vignette for more details).

#### Value

SpatRaster that includes a raster layer of genetic diversity and a raster layer of the number of samples within the window for each cell

#### Examples

```
load_mini_ex()
cpi <- circle_gd(mini_vcf, mini_coords, mini_lyr, fact = 2, maxdist = 5)</pre>
```

circle\_general *General function for making circular moving window maps* 

#### Description

Generate a continuous raster map using circular moving windows. While resist\_gd is built specifically for making maps of genetic diversity from vcfs,circle\_general can be used to make maps from different data inputs. Unlike resist\_gd, resist\_general will not convert your data into the correct format for calculations of different diversity metrics. See details for how to format data inputs for different statistics.

#### Usage

```
circle_general(
    x,
    coords,
    lyr,
    maxdist,
    distmat = NULL,
    stat,
    fact = 0,
    rarify = FALSE,
    rarify_nt = 2,
    rarify_nit = 5,
    min_n = 2,
    fun = mean,
```

```
L = NULL,
rarify_alleles = TRUE,
sig = 0.05,
...
```

x	data to be summarized by the moving window ( <i>note:</i> order matters! coords should be in the same order, there are currently no checks for this). The class of x required depends on the statistic being calculated (see the stat argument and the function description for more details)
coords	coordinates of samples as sf points, a two-column matrix, or a data.frame repre- senting x and y coordinates (see Details for important information about projec- tions)
lyr	SpatRaster or RasterLayer to slide the window across (see Details for important information about projections)
maxdist	maximum geographic distance used to define neighborhood; any samples further than this distance will not be included (this can be thought of as the neighborhood radius) Can either be (1) a single numeric value or (2) a SpatRaster where each pixel is the maximum distance to be used for that cell on the landscape (must be the same spatial scale as lyr).
distmat	distance matrix output from get_geodist (optional; can be used to save time on distance calculations)
stat	moving window statistic to calculate (see details). stat can generally be set to any function that will take xas input and return a single numeric value (for example, x can be a vector and stat can be set equal to a summary statistic like mean, sum, or sd)
fact	aggregation factor to apply to lyr (defaults to 0; <i>note:</i> increasing this value reduces computational time)
rarify	if rarify = TRUE, rarefaction is performed (defaults to FALSE)
rarify_n	if rarify = TRUE, number of points to use for rarefaction (defaults to min_n)
rarify_nit	if rarify = TRUE, number of iterations to use for rarefaction (defaults to 5). Can also be set to "all" to use all possible combinations of samples of size rarify_n within the window.
min_n	minimum number of samples to use in calculations (any focal cell with a window containing less than this number of samples will be assigned a value of NA; defaults to 2)
fun	function to use to summarize rarefaction results (defaults to mean, must take na.rm = TRUE as an argument)
L	for calculating "pi", L argument in pi.dosage function. Return the average nucleotide diversity per nucleotide given the length L of the sequence. The wingen default is $L =$ "nvariants" which sets L to the number of variants in the VCF. If L = NULL, returns the sum over SNPs of nucleotide diversity ( <i>note:</i> L = NULL is the pi.dosage default which wingen does not use)

rarify_alleles	for calculating "biallelic_richness", whether to perform rarefaction of al- lele counts as in allelic.richness (defaults to TRUE)
sig	for calculating "hwe", significance threshold (i.e., alpha level) to use for hardy-weinberg equilibrium tests (defaults to $0.05$ )
	if a function is provided for stat, additional arguments to pass to the stat function (e.g. if stat = mean, users may want to set na.rm = TRUE)

#### Details

To calculate genetic diversity statistics with the built in wingen functions, data must be formatted as such:

- for "pi" or "biallelic\_richness", x must be a dosage matrix with values of 0, 1, or 2
- for "Ho", x must be a heterozygosity matrix where values of 0 = homozygosity and values of 1 = heterozygosity
- for "allelic\_richness" or "hwe, x must be a genind type object
- for "basic\_stats", x must be a hierfstat type object

Otherwise, stat can be any function that takes a matrix or data frame and outputs a single numeric value (e.g., a function that produces a custom diversity index); however, this should be attempted with caution since this functionality has not have been tested extensively and may produce errors.

#### Value

SpatRaster that includes a raster layer of genetic diversity and a raster layer of the number of samples within the window for each cell

coords\_to\_raster Create a raster from coordinates

#### Description

Generate a raster layer from coordinates which can be used in window\_gd as the RasterLayer to move the window across

#### Usage

```
coords_to_raster(
   coords,
   buffer = 0,
   res = 1,
   agg = NULL,
   disagg = NULL,
   plot = FALSE
)
```

#### get\_geodist

## Arguments

coords	coordinates of samples as sf points, a SpatVector, a two-column matrix, or a data.frame with x and y coordinates
buffer	size of buffer to add to edge of raster (defaults to 0)
res	desired resolution of raster (defaults to 1). Can be a single value for square cells or a vector with two values representing x and y resolutions
agg	aggregation factor to apply to raster (defaults to NULL)
disagg	disaggregation factor to apply to raster (defaults to NULL)
plot	whether to plot resulting raster with coords (defaults to FALSE)

#### Value

RasterLayer

#### Examples

```
load_mini_ex()
coords_to_raster(mini_coords, buffer = 1, plot = TRUE)
```

get\_geodist

Get a matrix of geographic distances for circle\_gd

#### Description

Create a distance matrix based on coordinates and a raster layer. The output is a distance matrix where rows represent cells on the landscape and columns represent individual locations on the landscape. Each value is the geographic distance between each individual and each cell calculated using st\_distance. This matrix is used by circle\_gd. If coords\_only = TRUE, the result is a distance matrix for the sample coordinates only.

#### Usage

```
get_geodist(coords, lyr = NULL, fact = 0, coords_only = FALSE)
```

coords	coordinates of samples as sf points, a two-column matrix, or a data.frame repre- senting x and y coordinates (see Details for important information about projec- tions)
lyr	SpatRaster or RasterLayer for generating distances (not required if coords_only = TRUE)
fact	aggregation factor to apply to lyr (defaults to 0; <i>note:</i> increasing this value reduces computational time)
coords_only	whether to return distances only for sample coordinates

## Value

a distance matrix used by circle\_gd

#### Examples

```
load_mini_ex()
distmat <- get_geodist(mini_coords, mini_lyr)</pre>
```

get\_resdist

```
Get a matrix of resistance distances for resist_gd
```

## Description

Create a distance matrix based on coordinates and a connectivity layer. The output is a distance matrix where rows represent cells on the landscape and columns represent individual locations on the landscape. Each value is the resistance distance between each sample and each cell calculated using the gdistance package. This matrix is used by resist\_gd. If coords\_only = TRUE, the result is a distance matrix for the sample coordinates only.

## Usage

```
get_resdist(
  coords,
  lyr,
  fact = 0,
  transitionFunction = mean,
  directions = 8,
  geoCorrection = TRUE,
  coords_only = FALSE
)
```

coords	coordinates of samples as sf points, a two-column matrix, or a data.frame repre- senting x and y coordinates (see Details for important information about projec- tions)	
lyr	conductivity layer (higher values should mean greater conductivity) for generat- ing distances. Can be either a SpatRaster or RasterLayer.	
fact	aggregation factor to apply to lyr (defaults to 0; <i>note:</i> increasing this value reduces computational time)	
transitionFunction		
	function to calculate transition values from grid values (defaults to mean)	
directions	directions in which cells are connected (4, 8, 16, or other), see adjacent (defaults to 8)	

## ggplot\_count

geoCorrection	whether to apply correction to account for local distances (defaults to TRUE). Geographic correction is necessary for all objects of the class Transition that are either: (1) based on a grid in a geographic (lonlat) projection and covering a large area; (2) made with directions > 4 (see geoCorrection for more details).
coords_only	whether to return distances only for sample coordinates

## Value

a distance matrix used by resist\_gd

## Examples

```
load_mini_ex()
distmat <- get_resdist(mini_coords, mini_lyr)</pre>
```

ggplot\_count

Plot moving window map of sample counts

## Description

Plot sample counts layer produced by window\_gd or krig\_gd

## Usage

```
ggplot_count(x, index = NULL, col = viridis::mako(100))
```

## Arguments

Х	single SpatRaster of counts or SpatRaster where indexed layer is sample counts
index	index of raster layers to plot (defaults to plotting the one called "sample_count", if more than one layer is provided)
col	color palette to use for plotting (defaults to viridis::mako palette)

## Value

list of ggplots

## Examples

```
data("mini_lyr")
ggplot_count(mini_lyr)
```

ggplot\_gd

## Description

Plot genetic diversity layer produced by window\_gd or krig\_gd

## Usage

```
ggplot_gd(x, bkg = NULL, index = NULL, col = viridis::magma(100))
```

## Arguments

х	output from window_gd or krig_gd (RasterStack where first layer is genetic diversity)
bkg	optional raster or sf polygon
index	index of raster layers to plot (defaults to plotting all of the layers except the one called "sample_count", if more than one layer is provided)
col	color palette to use for plotting (defaults to magma palette)

#### Value

list of ggplots

## Examples

```
data("mini_lyr")
ggplot_gd(mini_lyr)
```

krig\_gd

Krige moving window maps

## Description

Perform interpolation of the raster(s) produced by window\_gd using autoKrige

krig\_gd

## Usage

```
krig_gd(
 r,
 grd = NULL,
  index = 1,
  coords = NULL,
  agg_grd = NULL,
  disagg_grd = NULL,
  agg_r = NULL,
  disagg_r = NULL,
  autoKrige_output = FALSE,
 lower_bound = TRUE,
  upper_bound = TRUE,
 krig_method = "ordinary",
  resample = FALSE,
 resample_first = TRUE
)
```

r	SpatRaster produced by window_gd
grd	object to create grid for kriging; can be a SpatRaster or RasterLayer. If undefined, will use $r$ to create a grid.
index	integer indices of layers in raster stack to krige (defaults to 1; i.e., the first layer)
coords	if provided, kriging will occur based only on values at these coordinates. Can be provided as an sf points, a two-column matrix, or a data.frame representing x and y coordinates
agg_grd	factor to use for aggregation of grd, if provided (this will decrease the resolution of the final kriged raster; defaults to NULL)
disagg_grd	factor to use for disaggregation of grd, if provided (this will increase the resolution of the final kriged raster; defaults to NULL)
agg_r	factor to use for aggregation of r, if provided (this will decrease the number of points used in the kriging model; defaults to NULL)
disagg_r	factor to use for disaggregation, of r if provided (this will increase the number of points used in the kriging model; defaults to NULL)
autoKrige_outpu	t
	whether to return full output from autoKrige including uncertainty rasters (de- faults to FALSE). If TRUE, returns a list with the kriged input raster layer ("raster"), kriged variance ("var"), kriged standard deviation ("stdev"), and full autoKrige output ("autoKrige_output").
lower_bound	if TRUE (default), converts all values in the kriged raster less than the minimum value of the input raster, to that minimum
upper_bound	if TRUE (default), converts all values in the kriged raster greater than the maximum value of the input raster, to that maximum

krig_method	method to use for kriging. If ordinary, ordinary/simple kriging is performed (formula: ~ 1; default). If universal, universal kriging is performed (formula $= x + y$ ).
resample	whether to resample $grd$ or $r$ . Set to " $r$ " to resample $r$ to $grd$ . Set to " $grd$ " to resample $grd$ to $r$ (defaults to FALSE for no resampling)
resample_first	if aggregation or disaggregation is used in addition to resampling, specifies whether to resample before (resample_first = TRUE) or after (resample_first = FALSE) aggregation/disaggregation (defaults to TRUE)

#### Value

a SpatRaster object or a list of autoKrige outputs (if autoKrige\_output = TRUE)

## Examples

```
load_mini_ex()
wpi <- window_gd(mini_vcf, mini_coords, mini_lyr, L = 10, rarify = TRUE)
kpi <- krig_gd(wpi, mini_lyr)
plot_gd(kpi, main = "Kriged Pi")</pre>
```

load\_middle\_earth\_ex Middle earth example

## Description

Loads middle earth example data

## Usage

```
load_middle_earth_ex(quiet = FALSE)
```

## Arguments

quiet whether to hide message (defaults to FALSE)

## Value

three objects are loaded (lotr\_vcf, lotr\_coords, and lotr\_lyrs)

## Examples

load\_middle\_earth\_ex()

load\_mini\_ex

## Description

Loads mini middle earth example data

## Usage

load\_mini\_ex(quiet = FALSE)

## Arguments

quiet

whether to hide message (defaults to FALSE)

#### Value

three objects are assigned in the GlobalEnv (vcf, coords, and lyr)

## Examples

load\_mini\_ex()

lotr\_coords Middle earth example coordinates

## Description

Middle earth example coordinates

#### Usage

lotr\_coords

#### Format

A data frame with 100 rows and 2 columns

- x x coordinate
- y y coordinate

#### Source

created from simulations in Bishop et al. (2023)

lotr\_lyr

## Description

RasterLayer of middle earth based on an example digital elevation model of Tolkien's Middle Earth produced by the Center for Geospatial Analysis at William & Mary (Robert, 2020).

## Usage

lotr\_lyr

## Format

RasterLayer

## Source

created from simulations in Bishop et al. (2023) based on Rose, Robert A. (2020) GIS & Middle Earth Presentation & Data Set. William & Mary. https://doi.org/10.21220/RKEZ-X707

lotr\_range

Middle earth example range polygon

## Description

sf polygon of range map

#### Usage

lotr\_range

#### Format

 $\mathbf{sf}$ 

#### Source

created from simulations in Bishop et al. (2023)

lotr\_vcf

### Description

A Variant Call Format data set

## Usage

lotr\_vcf

## Format

Object of class vcfR with 100 individuals and 1000 loci

#### Source

created from simulations in Bishop et al. (2023)

mask_gd	Mask moving window maps	
---------	-------------------------	--

## Description

Mask genetic diversity layer produced by window\_gd or krig\_gd

## Usage

mask\_gd(x, y, minval = NULL, maxval = NULL)

## Arguments

х	Raster object to mask
у	Raster object or Spatial object to use as mask
minval	if y is a Raster object, value of y below which to mask
maxval	if y is a Raster object, value of y above which to mask

## Value

RasterLayer

## Examples

```
data("mini_lyr")
mpi <- mask_gd(mini_lyr, mini_lyr, minval = 0.01)</pre>
```

mini\_coords

## Description

Mini middle earth example coordinates

## Usage

mini\_coords

#### Format

A data frame with 10 rows and 2 columns

x x coordinate

y y coordinate

## Source

created from simulations in Bishop et al. (2023)

mini\_lyr

Mini middle earth example raster

## Description

Small RasterLayer of middle earth based on an example digital elevation model of Tolkien's Middle Earth produced by the Center for Geospatial Analysis at William & Mary (Robert, 2020).

## Usage

mini\_lyr

#### Format

A RasterLayer of middle earth

#### Source

created from simulations in Bishop et al. (2023)

mini\_vcf

## Description

A Variant Call Format data set

## Usage

mini\_vcf

## Format

Object of class vcfR with 10 individuals and 10 loci

## Source

created from simulations in Bishop et al. (2023)

mini\_vcf\_NA Mini middle earth example vcf with NA values

## Description

A Variant Call Format data set with NA values

## Usage

mini\_vcf\_NA

## Format

Object of class vcfR with 10 individuals and 10 loci

#### Source

created from simulations in Bishop et al. (2023)

plot\_count

## Description

Plot sample counts layer produced by window\_gd or krig\_gd

## Usage

```
plot_count(
    x,
    index = NULL,
    breaks = 100,
    col = viridis::mako(breaks),
    main = NULL,
    box = FALSE,
    range = NULL,
    legend = TRUE,
    ...
)
```

## Arguments

х	single SpatRaster of counts or SpatRaster where indexed layer is sample counts
index	if a raster stack is provided, index of the sample count layer to plot (defaults to plotting the layer named "sample_count" or the last layer of the stack)
breaks	number of breaks to use in color scale (defaults to 10)
col	color palette to use for plotting (defaults to viridis::magma palette)
main	character. Main plot titles (one for each layer to be plotted). You can use arguments cex.main, font.main, col.main to change the appearance; and loc.main to change the location of the main title (either two coordinates, or a character value such as "topleft")
box	whether to include a box around the raster plot (defaults to FALSE)
range	numeric. minimum and maximum values to be used for the continuous legend
legend	whether to include legend
	arguments passed to plot("SpatRaster", "numeric") and additional graphical arguments

## Value

plot of sample counts

## Examples

```
data("mini_lyr")
plot_count(mini_lyr)
```

plot\_gd

## Description

Plot genetic diversity layer produced by window\_gd or krig\_gd

## Usage

```
plot_gd(
    x,
    bkg = NULL,
    index = NULL,
    col = viridis::magma(breaks),
    breaks = 100,
    main = NULL,
    box = FALSE,
    range = NULL,
    legend = TRUE,
    ...
)
```

## Arguments

x	output from window_gd or krig_gd (SpatRaster where first layer is genetic diversity)
bkg	optional SpatRaster or other spatial object that will be plotted as the "back-ground" in gray
index	if a raster stack is provided, index of the layer to plot (defaults to plotting all layers except layers named "sample_count")
col	color palette to use for plotting (defaults to magma palette)
breaks	number of breaks to use in color scale (defaults to 100)
main	character. Main plot titles (one for each layer to be plotted). You can use arguments cex.main, font.main, col.main to change the appearance; and loc.main to change the location of the main title (either two coordinates, or a character value such as "topleft")
box	whether to include a box around the Raster plot (defaults to FALSE)
range	numeric. minimum and maximum values to be used for the continuous legend
legend	whether to include legend
	arguments passed to plot("SpatRaster", "numeric") and additional graphical arguments

## Value

plot of genetic diversity

## Examples

```
data("mini_lyr")
plot_gd(mini_lyr)
```

preview\_gd

Preview moving window and sample counts

## Description

Generate a preview of moving window size and sample counts based on the coordinates and parameters to be supplied to window\_gd, circle\_gd, or resist\_gd. The method to be used should be specified with method = "window", "circle", or "resist". For method = "window", wdim must be specified. For method = "circle" or "resist", maxdist must be specified and distmat can also optionally be specified.

#### Usage

```
preview_gd(
  lyr,
  coords,
  method = "window",
  wdim = 3,
  maxdist = NULL,
  distmat = NULL,
  fact = 0,
  sample_count = TRUE,
  min_n = 0,
  plot = TRUE
)
```

#### Arguments

lyr	SpatRaster or RasterLayer to slide the window across (see Details for important information about projections). For method = "resist" this should also be the conductivity layer (see resist_gd)
coords	coordinates of samples as sf points, a two-column matrix, or a data.frame repre- senting x and y coordinates (see Details for important information about projec- tions)
method	which method to use to create preview ("window" for window_gd, "circle" for circle_gd, or "resist" for resist_gd; defaults to "window")
wdim	if method = "window", dimensions (height x width) of window; if only one value is provided, a square window is created (defaults to 3 x 3 window)
maxdist	if method = "circle" or method = "resist, the maximum geographic distance used to define the neighborhood; any samples further than this distance will not be included (see get_geodist or get_resdist)

20

#### resist\_gd

distmat	if method = "circle" or method = "resist", an optional distance matrix to be used output from either get_geodist or get_resdist, respectively. If not provided, one will be automatically calculated.
fact	aggregation factor to apply to lyr (defaults to 0; <i>note:</i> increasing this value reduces computational time)
sample_count	whether to create plot of sample counts for each cell (defaults to TRUE)
min_n	minimum number of samples to use in calculations (any focal cell with a window containing less than this number of samples will be assigned a value of NA)
plot	whether to plot results (default = TRUE)

## Details

Coordinates and rasters should be in a projected (planar) coordinate system such that raster cells are of equal sizes. Therefore, spherical systems (including latitute-longitude coordinate systems) should be projected prior to use. Transformation can be performed using st\_set\_crs for coordinates or project for rasters (see vignette for more details).

## Value

Plots preview of window and returns SpatRaster with sample counts layer (if sample\_count = TRUE)

## Examples

```
load_mini_ex()
preview_gd(mini_lyr, mini_coords, wdim = 3, fact = 3, sample_count = TRUE, min_n = 2)
```

resist\_gd

Create a moving window map of genetic diversity based on resistance

#### Description

Generate a continuous raster map of genetic diversity using resistance distances calculated with a conductivity surface

#### Usage

```
resist_gd(
  gen,
  coords,
  lyr,
  maxdist,
  distmat = NULL,
  stat = "pi",
  fact = 0,
  rarify = FALSE,
```

```
rarify_n = 2,
rarify_nit = 5,
min_n = 2,
fun = mean,
L = "nvariants",
rarify_alleles = TRUE,
sig = 0.05,
transitionFunction = mean,
directions = 8,
geoCorrection = TRUE
)
```

## Arguments

gen	genetic data either as an object of type vcf or a path to a vcf file ( <i>note:</i> order matters! The coordinate and genetic data should be in the same order; there are currently no checks for this)
coords	coordinates of samples as sf points, a two-column matrix, or a data.frame repre- senting x and y coordinates (see Details for important information about projec- tions)
lyr	conductivity layer (higher values should mean greater conductivity) to move window across. Can be either a SpatRaster or RasterLayer.
maxdist	maximum cost distance used to define neighborhood; any samples further than this cost distance will not be included (this can be thought of as the neighbor- hood radius, but in terms of cost distance). Can either be (1) a single numeric value or (2) a SpatRaster where each pixel is the maximum distance to be used for that cell on the landscape (must be the same spatial scale as lyr).
distmat	distance matrix output from get_resdist (optional; can be used to save time on distance calculations)
stat	genetic diversity statistic(s) to calculate (see Details, defaults to "pi"). Can be a single statistic or a vector of statistics
fact	aggregation factor to apply to lyr (defaults to 0; <i>note:</i> increasing this value reduces computational time)
rarify	if rarify = TRUE, rarefaction is performed (defaults to FALSE)
rarify_n	if rarify = TRUE, number of points to use for rarefaction (defaults to min_n)
rarify_nit	if rarify = TRUE, number of iterations to use for rarefaction (defaults to 5). Can also be set to "all" to use all possible combinations of samples of size rarify_n within the window.
min_n	minimum number of samples to use in calculations (any focal cell with a window containing less than this number of samples will be assigned a value of NA; defaults to 2)
fun	function to use to summarize rarefaction results (defaults to mean, must take na.rm = TRUE as an argument)
L	for calculating "pi", L argument in pi.dosage function. Return the average nucleotide diversity per nucleotide given the length L of the sequence. The wingen

22

		default is $L =$ "nvariants" which sets L to the number of variants in the VCF. If $L = NULL$ , returns the sum over SNPs of nucleotide diversity ( <i>note:</i> $L = NULL$ is the pi.dosage default which wingen does not use)
	rarify_alleles	for calculating "biallelic_richness", whether to perform rarefaction of al- lele counts as in allelic.richness (defaults to TRUE)
	sig	for calculating "hwe", significance threshold (i.e., alpha level) to use for hardy- weinberg equilibrium tests (defaults to 0.05)
transitionFunction		
		function to calculate transition values from grid values (defaults to mean)
	directions	directions in which cells are connected (4, 8, 16, or other), see adjacent (defaults to 8)
	geoCorrection	whether to apply correction to account for local distances (defaults to TRUE). Geographic correction is necessary for all objects of the class Transition that are either: (1) based on a grid in a geographic (lonlat) projection and covering a large area; (2) made with directions > 4 (see geoCorrection for more details).

#### Details

Coordinates and rasters should be in a Euclidean coordinate system (i.e., UTM coordinates) such that raster cell width and height are equal distances. As such, longitude-latitude systems should be transformed before using dist\_gd. Transformation can be performed using st\_set\_crs for coordinates or project for rasters (see vignette for more details).

## Value

SpatRaster that includes a raster layer of genetic diversity and a raster layer of the number of samples within the window for each cell

#### Examples

```
load_mini_ex()
rpi <- resist_gd(mini_vcf, mini_coords, mini_lyr, maxdist = 50)</pre>
```

resist\_general General function for making resistance-based maps

#### Description

Generate a continuous raster map using resistance distances. While resist\_gd is built specifically for making maps of genetic diversity from vcfs,resist\_general can be used to make maps from different data inputs. Unlike resist\_gd, resist\_general will not convert your data into the correct format for calculations of different diversity metrics. See details for how to format data inputs for different statistics.

## Usage

```
resist_general(
 х,
 coords,
 lyr,
 maxdist,
 distmat = NULL,
 stat,
 fact = 0,
 rarify = FALSE,
 rarify_n = 2,
 rarify_nit = 5,
 min_n = 2,
  fun = mean,
 L = NULL,
  rarify_alleles = TRUE,
  sig = 0.05,
  transitionFunction = mean,
 directions = 8,
 geoCorrection = TRUE,
  . . .
)
```

## Arguments

x	data to be summarized by the moving window ( <i>note:</i> order matters! coords should be in the same order, there are currently no checks for this). The class of x required depends on the statistic being calculated (see the stat argument and the function description for more details)
coords	coordinates of samples as sf points, a two-column matrix, or a data.frame repre- senting x and y coordinates (see Details for important information about projec- tions)
lyr	SpatRaster or RasterLayer to slide the window across (see Details for important information about projections)
maxdist	maximum cost distance used to define neighborhood; any samples further than this cost distance will not be included (this can be thought of as the neighbor- hood radius, but in terms of cost distance). Can either be (1) a single numeric value or (2) a SpatRaster where each pixel is the maximum distance to be used for that cell on the landscape (must be the same spatial scale as lyr).
distmat	distance matrix output from get_resdist (optional; can be used to save time on distance calculations)
stat	moving window statistic to calculate (see details). stat can generally be set to any function that will take xas input and return a single numeric value (for example, x can be a vector and stat can be set equal to a summary statistic like mean, sum, or sd)
fact	aggregation factor to apply to lyr (defaults to 0; <i>note:</i> increasing this value reduces computational time)

24

rarify	if rarify = TRUE, rarefaction is performed (defaults to FALSE)
rarify_n	if rarify = TRUE, number of points to use for rarefaction (defaults to min_n)
rarify_nit	if rarify = TRUE, number of iterations to use for rarefaction (defaults to 5). Can also be set to "all" to use all possible combinations of samples of size rarify_n within the window.
min_n	minimum number of samples to use in calculations (any focal cell with a window containing less than this number of samples will be assigned a value of NA; defaults to 2)
fun	function to use to summarize rarefaction results (defaults to mean, must take na.rm = TRUE as an argument)
L	for calculating "pi", L argument in pi.dosage function. Return the average nucleotide diversity per nucleotide given the length L of the sequence. The wingen default is $L =$ "nvariants" which sets L to the number of variants in the VCF. If L = NULL, returns the sum over SNPs of nucleotide diversity ( <i>note:</i> L = NULL is the pi.dosage default which wingen does not use)
rarify_alleles	for calculating "biallelic_richness", whether to perform rarefaction of al- lele counts as in allelic.richness (defaults to TRUE)
sig	for calculating "hwe", significance threshold (i.e., alpha level) to use for hardy- weinberg equilibrium tests (defaults to 0.05)
transitionFunc	tion
	function to calculate transition values from grid values (defaults to mean)
directions	directions in which cells are connected (4, 8, 16, or other), see adjacent (defaults to 8)
geoCorrection	whether to apply correction to account for local distances (defaults to TRUE). Geographic correction is necessary for all objects of the class Transition that are either: (1) based on a grid in a geographic (lonlat) projection and covering a large area; (2) made with directions > 4 (see geoCorrection for more details).
	if a function is provided for stat, additional arguments to pass to the stat function (e.g. if stat = mean, users may want to set na.rm = TRUE)

#### Details

To calculate genetic diversity statistics with the built in wingen functions, data must be formatted as such:

- for "pi" or "biallelic\_richness", x must be a dosage matrix with values of 0, 1, or 2
- for "Ho", x must be a heterozygosity matrix where values of 0 = homozygosity and values of 1 = heterozygosity
- for "allelic\_richness" or "hwe, x must be a genind type object
- for "basic\_stats", x must be a hierfstat type object

Otherwise, stat can be any function that takes a matrix or data frame and outputs a single numeric value (e.g., a function that produces a custom diversity index); however, this should be attempted with caution since this functionality has not have been tested extensively and may produce errors.

## Value

SpatRaster that includes a raster layer of genetic diversity and a raster layer of the number of samples within the window for each cell

vcf\_to\_dosage Convert a vcf to a dosage matrix

## Description

Convert a vcf to a dosage matrix

#### Usage

vcf\_to\_dosage(x)

#### Arguments

Х

can either be an object of class 'vcfR' or a path to a .vcf file

#### Value

dosage matrix

window\_gd

Create a moving window map of genetic diversity

## Description

Generate a continuous raster map of genetic diversity using moving windows.

#### Usage

```
window_gd(
  gen,
  coords,
  lyr,
  stat = "pi",
  wdim = 3,
  fact = 0,
  rarify = FALSE,
  rarify_n = NULL,
  rarify_nit = 5,
  min_n = 2,
  fun = mean,
  L = "nvariants",
```

## window\_gd

```
rarify_alleles = TRUE,
sig = 0.05,
crop_edges = FALSE,
...
```

gen	genetic data either as an object of type vcf or a path to a vcf file ( <i>note:</i> order matters! The coordinate and genetic data should be in the same order; there are currently no checks for this)
coords	coordinates of samples as sf points, a two-column matrix, or a data.frame repre- senting x and y coordinates (see Details for important information about projec- tions)
lyr	SpatRaster or RasterLayer to slide the window across (see Details for important information about projections)
stat	genetic diversity statistic(s) to calculate (see Details, defaults to "pi"). Can be a single statistic or a vector of statistics
wdim	dimensions (height x width) of window; if only one value is provided, a square window is created (defaults to 3 x 3 window)
fact	aggregation factor to apply to lyr (defaults to 0; <i>note:</i> increasing this value reduces computational time)
rarify	if rarify = TRUE, rarefaction is performed (defaults to FALSE)
rarify_n	if rarify = TRUE, number of points to use for rarefaction (defaults to min_n)
rarify_nit	if rarify = TRUE, number of iterations to use for rarefaction (defaults to 5). Can also be set to "all" to use all possible combinations of samples of size rarify_n within the window.
min_n	minimum number of samples to use in calculations (any focal cell with a window containing less than this number of samples will be assigned a value of NA; defaults to 2)
fun	function to use to summarize rarefaction results (defaults to mean, must take na.rm = TRUE as an argument)
L	for calculating "pi", L argument in pi.dosage function. Return the average nucleotide diversity per nucleotide given the length L of the sequence. The wingen default is $L =$ "nvariants" which sets L to the number of variants in the VCF. If L = NULL, returns the sum over SNPs of nucleotide diversity ( <i>note:</i> L = NULL is the pi.dosage default which wingen does not use)
rarify_alleles	for calculating "biallelic_richness", whether to perform rarefaction of al- lele counts as in allelic.richness (defaults to TRUE)
sig	for calculating "hwe", significance threshold (i.e., alpha level) to use for hardy- weinberg equilibrium tests (defaults to 0.05)
crop_edges	whether to remove cells on the edge of the raster where the window is incomplete (defaults to FALSE)

deprecated this was intended to be used to pass additional arguments to the stat function, however now formal arguments are used instead (see L, rarify\_alleles, and sig). Passing additional arguments using ... is still possible with the \*\_general() functions.

#### Details

. . .

Coordinates and rasters should be in a projected (planar) coordinate system such that raster cells are of equal sizes. Therefore, spherical systems (including latitute-longitude coordinate systems) should be projected prior to use. Transformation can be performed using st\_set\_crs for coordinates or project for rasters (see vignette for more details).

Current genetic diversity metrics that can be specified with stat include:

- "pi" for nucleotide diversity (default) calculated using hierfstat pi.dosage
- "Ho" for average observed heterozygosity across all sites
- "allelic\_richness" for average number of alleles across all sites
- "biallelic\_richness" for average allelic richness across all sites for a biallelic dataset (this option is faster than "allelic\_richness")
- "hwe" for the proportion of sites that are not in Hardy–Weinberg equilibrium, calculated using pegas hw.test at the 0.05 level (other alpha levels can be specified by adding the sig argument; e.g., sig = 0.10).
- "basic\_stats" for a series of statistics produced by hierfstat basic.stats including mean observed heterozygosity (same as Ho), mean gene diversities within population (Hs), Gene diversities overall (Ht), and Fis following Nei (1987). Population-based statistics (e.g., FST) normally reported by basic.stats are not included as they are not meaningful within the individualbased moving windows.

#### Value

SpatRaster that includes raster layers of genetic diversity and a raster layer of the number of samples within the window for each cell

## Examples

```
load_mini_ex()
wpi <- window_gd(mini_vcf, mini_coords, mini_lyr, rarify = TRUE)</pre>
```

window\_general General function for making moving window maps

#### Description

Generate a continuous raster map using moving windows. While window\_gd is built specifically for making moving window maps of genetic diversity from vcfs, window\_general can be used to make moving window maps from different data inputs. See details for how to format data inputs for different statistics.

window\_general

## Usage

```
window_general(
 х,
 coords,
 lyr,
  stat,
 wdim = 3,
 fact = 0,
 rarify = FALSE,
 rarify_n = NULL,
 rarify_nit = 5,
 min_n = 2,
 fun = mean,
 L = "nvariants",
 rarify_alleles = TRUE,
 sig = 0.05,
 crop_edges = FALSE,
  . . .
)
```

x	data to be summarized by the moving window ( <i>note:</i> order matters! coords should be in the same order, there are currently no checks for this). The class of x required depends on the statistic being calculated (see the stat argument and the function description for more details)
coords	coordinates of samples as sf points, a two-column matrix, or a data.frame repre- senting x and y coordinates (see Details for important information about projec- tions)
lyr	SpatRaster or RasterLayer to slide the window across (see Details for important information about projections)
stat	moving window statistic to calculate (can either be "pi" for nucleotide di- versity (x must be a dosage matrix), "Ho" for average observed heterozygos- ity across all loci (x must be a heterozygosity matrix), "allelic_richness" for average allelic richness across all loci (x must be a genind type object), "biallelic_richness" to get average allelic richness across all loci for a bial- lelic dataset (x must be a dosage matrix). stat can also be set to any function that will take xas input and return a single numeric value (for example, x can be a vector and stat can be set equal to a summary statistic like mean, sum, or sd)
wdim	dimensions (height x width) of window; if only one value is provided, a square window is created (defaults to 3 x 3 window)
fact	aggregation factor to apply to lyr (defaults to 0; <i>note:</i> increasing this value reduces computational time)
rarify	if rarify = TRUE, rarefaction is performed (defaults to FALSE)
rarify_n	if rarify = TRUE, number of points to use for rarefaction (defaults to min_n)

rarify_nit	if rarify = TRUE, number of iterations to use for rarefaction (defaults to 5). Can also be set to "all" to use all possible combinations of samples of size rarify_n within the window.
min_n	minimum number of samples to use in calculations (any focal cell with a window containing less than this number of samples will be assigned a value of NA; defaults to 2)
fun	function to use to summarize rarefaction results (defaults to mean, must take na.rm = TRUE as an argument)
L	for calculating "pi", L argument in pi.dosage function. Return the average nucleotide diversity per nucleotide given the length L of the sequence. The wingen default is $L =$ "nvariants" which sets L to the number of variants in the VCF. If L = NULL, returns the sum over SNPs of nucleotide diversity ( <i>note:</i> L = NULL is the pi.dosage default which wingen does not use)
rarify_alleles	for calculating "biallelic_richness", whether to perform rarefaction of al- lele counts as in allelic.richness (defaults to TRUE)
sig	for calculating "hwe", significance threshold (i.e., alpha level) to use for hardy-weinberg equilibrium tests (defaults to $0.05$ )
crop_edges	whether to remove cells on the edge of the raster where the window is incomplete (defaults to FALSE)
	if a function is provided for stat, additional arguments to pass to the stat function (e.g. if stat = mean, users may want to set na.rm = TRUE)

## Details

To calculate genetic diversity statistics with the built in wingen functions, data must be formatted as such:

- for "pi" or "biallelic\_richness", x must be a dosage matrix with values of 0, 1, or 2
- for "Ho", x must be a heterozygosity matrix where values of 0 = homozygosity and values of 1 = heterozygosity
- for "allelic\_richness" or "hwe, x must be a genind type object
- for "basic\_stats", x must be a hierfstat type object

Otherwise, stat can be any function that takes a matrix or data frame and outputs a single numeric value (e.g., a function that produces a custom diversity index); however, this should be attempted with caution since this functionality has not have been tested extensively and may produce errors.

## Value

SpatRaster that includes a raster layer of genetic diversity and a raster layer of the number of samples within the window for each cell

# Index

\* datasets lotr\_coords, 13 lotr\_lyr, 14 lotr\_range, 14 lotr\_vcf, 15 mini\_coords, 16 mini\_lyr, 16 mini\_vcf, 17 mini\_vcf\_NA, 17 adjacent, 8, 23, 25 allelic.richness, 3, 6, 23, 25, 27, 30 autoKrige, 10-12 basic.stats, 28 circle\_gd, 2, 7, 8, 20 circle\_general, 4 coords\_to\_raster, 6 deprecated, 28 geoCorrection, 9, 23, 25 get\_geodist, 3, 5, 7, 20, 21 get\_resdist, 8, 20-22, 24 ggplot\_count, 9 ggplot\_gd, 10 hw.test, 28 krig\_gd, 9, 10, 10, 15, 18, 19 load\_middle\_earth\_ex, 12 load\_mini\_ex, 13 lotr\_coords, 13 lotr\_lyr, 14 lotr\_range, 14 lotr\_vcf, 15

magma, *10*, *19* mask\_gd, 15 mini\_coords, 16
mini\_lyr, 16
mini\_vcf, 17
mini\_vcf\_NA, 17

pi.dosage, 3, 5, 22, 23, 25, 27, 28, 30
plot\_count, 18
plot\_gd, 19
preview\_gd, 20
project, 4, 21, 23, 28

resist\_gd, 4, 8, 9, 20, 21, 23
resist\_general, 23

st\_distance, 7
st\_set\_crs, 4, 21, 23, 28

 $vcf_to_dosage, 26$ 

window\_gd, 6, 9–11, 15, 18–20, 26, 28 window\_general, 28